BLISS Gel DNA Extraction Kit (B110-001)

Kit contents and storage conditions

Contents*	Size	Storage temperature
Mini column and Collecting tube	200 ea	Room Temperature
GB buffer**	200 ml	
BW buffer (Concentrates***)	32 ml***	
EB buffer****	30 ml	

* All components of this kit should be stored at room temperature (15~25 °C). Long exposure to heat source can deteriorate the performance of kit significantly.

** During shipment or storage under cold ambient condition, a precipitate can be formed in GB buffer. Heat the bottle at 20°C ~40°C to dissolve completely in such a case.

*** BW buffer is provided as concentrates. Ethanol must be added before first use as the indication on the bottle label.

**** 10mM TrisCl, pH 8.5

Product use limitations

BLISS Gel DNA extraction kit is intended for research uses only. This kit is not intended for diagnosis or treatment for human. All due care and attention should be exercised in the handling of the products.

Safety information

BLISS Gel DNA extraction kit contains irritants which are harmful when in contact with skin or eyes, or inhaled or swallowed. Care should be taken when handling this product. Always wear gloves and eye protection, and follow standard safety precautions.

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Protocol for extraction of DNA from agarose gel.

- Prepare water bath or dry bath at 50°C
- Required consumables; 1.5 ml centrifuge tube
- Do not use the precipitated buffers. If a precipitate forms in a buffer, dissolve completely at 20°C ~40°C before use.
- Before first use, add *absolute* ethanol into a wash buffer as below.

Buffer name	Contents	Ethanol to be added	The final volume
BW buffer	32 ml	128 ml	160 ml

1. Excise the DNA band of interest using a sterile razor blade or scalpel on a transilluminator.

   Minimize a gel volume by cutting the gel slice as small as possible.

   Do cutting the gel rapidly on a long wave length-UV.

2. Weigh the gel slice in a micro centrifuge tube and add 3 volumes(ul) of GB buffer into the tube per 1 volume(mg) of sliced gel.

   Add 300ul of GB buffer into the tube for 100mg of sliced gel.

3. Incubate at 50°C until the agarose gel is completely melted.

   A sliced gel is generally melted completely in 10 mins.

4. Briefly vortex the tube for homogenization of the mixture.
5. Transfer 700 ul of the mixture into a spin column and centrifuge for 30 secs at 13,000 xg.
6. Remove the spin column, discard the pass-through, and insert the column back into the collection tube.

   If the mixtures are remained, repeat the step 5~6 with them.

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7. Apply700 ul of BW buffer into the column and centrifuge for 30 secs at 13,000 xg.
8. Remove the spin column, discard the pass-through, and insert the column back into the collection tube.
9. Centrifuge for 1 min at 13,000xg for drying the membrane and transfer the spin column into a new 1.5 ml tube.
10. Apply 50ul of EB buffer or sterile water to the center of spin column membrane and let it stand for 1 min.
11. Centrifuge at 13,000 xg for 1 min for eluting the DNA.