BLISS Plasmid DNA mini prep kit (B100-001)

Kit contents and storage conditions

Contents*	Size	Storage temperature
Mini column and Collecting tube	200 ea	Room Temperature
BP1 buffer	60 ml	
BP2** buffer	60 ml	
BP3** buffer	80 ml	
BPB buffer (Concentrates***)	(35mlx2) 70 ml***	
BW buffer (Concentrates***)	(14mlx2) 28 ml***	
EB buffer****	30 ml	
RNase A solution (20 mg/ml)	300 ul	

* All components of this kit should be stored at room temperature (15~25 °C). Long exposure to heat source can deteriorate the performance of kit significantly.

** During shipment or storage under cold ambient condition, a precipitate can be formed in BP 2 and/or3. Heat the bottle at 20°C ~40°C to dissolve completely in such a case.

*** Wash buffers are provided as concentrates. Ethanol must be added before first use as the indication on the bottle labels.

**** 10 mM TrisCl, pH 8.5

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Product use limitations

BLISS Plasmid mini prep kit is intended for research uses only. This kit is not intended for diagnosis or treatment for human. All due care and attention should be exercised in the handling of the products.

Safety information

BLISS Plasmid mini prep kit contains irritants which are harmful when in contact with skin or eyes, or inhaled or swallowed. Care should be taken when handling this product. Always wear gloves and eye protection, and follow standard safety precautions.

Protocol for preparation of plasmid DNA

- Required consumables; 1.5 ml centrifuge tube
- Do not use the precipitated buffers. If a precipitate forms in a buffer, dissolve completely at 20°C ~40°C before use.
- Before first use,

① add*absolute* ethanol into wash buffers as below.

Buffer name	Contents	Ethanol to be added	The final volume
BPB buffer	70 ml	70 ml	140 ml
BW buffer	28 ml	112 ml	140 ml

② Add all of RNase A solution into the bottle of BP1 and mix well by gentle swirling. Store the BP1at 4°C after addition of RNase A.

1. Harvest up to 5 ml of bacterial culture by centrifugation for 5 mins at 10,000 xg. Discard the supernatant as much as possible without disturbing the pellet.
2. Resuspend the pelleted cells thoroughly in 250 ul of BP1. Transfer the suspension to a new 1.5 ml microcentrifuge tube.
3. Add 250 ul of BP2 and mix by inverting the tube 4 times. Let it stand until the mixture become clear.

   Do NOT vortex at this step and do NOT incubate the mixture for more than 5 mins.
4. Add 350 ul of BP3 and immediately mix by inverting the tube 4~6 times.
5. Centrifuge for 10 mins at 13,000 xg.
6. Transfer carefully the supernatant to a spin column and centrifuge for 30 secs at 13,000 xg.
7. Remove the spin column, discard the pass-through, and insert the column back into the collection tube.
8. Apply600 ul of BPB into the column and centrifuge for 30 secs at 13,000 xg.
9. Remove the spin column, discard the pass-through, and insert the column back into the collection tube.
10. Apply 600 ul of BW into the column and centrifuge for 30 secs at 13,000 xg.
11. Remove the spin column, discard the pass-through, and insert the column back into the collection tube.
12. Centrifuge for 1 min at 13,000xg for drying the membrane and transfer the spin column into a new 1.5 ml tube.
13. Apply 50 ul of EB buffer or sterile waterto the center of spin column membrane and let it stand for 1 min.
14. Centrifuge at 13,000 xg for 1 min for eluting the DNA.

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